

Vascular Endothelial Growth Factor as an Angiogenic Marker in Malignant Astrocytoma and Oligodendroglioma: An Indian Scenario

RAMYA S VOKUDA¹, BHEEMANATHI HANUMAN SRINIVAS², VENKATESH S MADHUGIRI³, SURENDRA KUMAR VERMA⁴

ABSTRACT

Introduction: The role of Vascular Endothelial Growth Factor (VEGF) in angiogenesis has been extensively studied in gliomas, such as astrocytoma and oligodendrogliomas, worldwide. However, there is limited information available with regard to the Indian population.

Aim: To study, whether VEGF is expressed in the Indian population in a pattern similar to that in other population.

Materials and Methods: In this prospective study approved by the Institute Ethics Committee for Human Studies at Jawaharlal Institute of Post Graduate Medical Education and Research (JIPMER) the patients operated for glioma in 2014 and 2015 (n = 60) were included. Tumours were graded as per the World Health Organization (WHO) grading system. VEGF expression in various grades was analysed using immunohistochemistry.

Results: Of the 60 patients included in this study, 15 were Grade II- (diffuse astrocytomas – 12; oligodendrogliomas- 3), 15 were Grade III-(anaplastic astrocytomas- 2; anaplastic oligodendrogliomas – 13) and 30 were Grade IV-glioblastomas. For VEGF antibody staining, two patients (3.33%) showed negative results and 58 patients (96%) showed positive results. VEGF positivity was 100% in Grade II and III, while it was 93.3% (28/30) in Grade IV tumours (p=0.012).

Conclusion: The expression of VEGF was associated with the grade of tumour, which gradually increased from Grade II to Grade IV. We conclude that VEGF-regulated angiogenesis plays an important role in tumour progression of astrocytomas and oligodendrogliomas in the Indian population as observed worldwide.

Keywords: Angiogenesis, Astrocytic tumours, Glial tumours

INTRODUCTION

In adults, malignant astrocytic tumours are an aggressive form of Central Nervous System (CNS) tumours, glioblastoma multiforme being the most common among them [1]. Patients with these tumours have a poor prognosis and low survival rates [2]. Despite advances in surgical procedures and radiation and chemotherapeutic regimens, these tumours are still a cause for high mortality.

VEGF is a potent pro-angiogenic cytokine upregulated in malignant glial tumours [2,3]. In glioblastoma, angiogenesis is mostly driven by hypoxia dependent and independent mechanisms [3-7]. The proangiogenic role of VEGF in glial tumours has been well documented in other countries [1,3]. These studies demonstrated the upregulation of VEGF, thus, anti-VEGF modalities and therapies have been developed. In the Indian population, however, this association has not been evaluated. In this study, we analysed the expression of VEGF in malignant astrocytomas and oligodendroglioma and determined whether the expression pattern in our population is similar to that observed in other studies.

MATERIALS AND METHODS

This is a prospective study approved by the Institute Ethics Committee for Human Studies at JIPMER. We obtained formalin fixed, paraffin embedded glial tumour biopsies {astrocytoma and oligodendroglioma (not otherwise specified)} from 60 patients (55% female; age range: 18–67 years (mean = 38.1 years) during a period of two years (2014 and 2015) from the Department of Neurosurgery, JIPMER, Puducherry, India. Informed consent was obtained from each patient.

All biopsies were characterised for tumour type and grade, according to the parameters established by the latest World Health Organization classification of brain tumours [8]. Of the 60 cases, 15 (25%) were classified as Grade II (diffuse astrocytoma-12; oligodendroglioma – 3), 15 (25%) as Grade III (anaplastic

astrocytoma – 2; anaplastic oligodendroglioma – 13) and 30 (50%) as Grade IV (glioblastoma).

Grade I tumours were excluded from the study as they are circumscribed benign tumours, in contrast to other gliomas which are mostly infiltrating and have tendency towards progression to malignancy. Also, the grade I tumours follow a distinctive molecular pathway unlike their malignant counter parts.

Immunohistochemistry: VEGF expression was analysed using immunohistochemistry. Sections were deparaffinised in xylene and then washed with alcohol; endogenous peroxidase was quenched using 3% H₂O₂ in methanol for 10 mins. The sections were then rehydrated in distilled water, and the slides were treated for nonenzymatic antigen retrieval in citrate buffer, pH 6.0 at 110 °C for one hour. They were then incubated with anti-VEGF primary antibody (rabbit polyclonal antibody prediluted in Phosphate Buffered Saline (PBS); BioGenex, San Ramen, CA, USA at room temperature (23 °C) for one hour in a humidified chamber. The slides were rinsed with Tris buffer and incubated with secondary antibody (Dako, Carpinteria, CA, USA) for 40 mins. They were rinsed again with Tris buffer and then incubated with Diaminobenzidine (DAB) for 15 mins. They were counter stained with haematoxylin, and then dried and mounted with di-n-butyl phthalate in xylene.

Scoring system: We used the scoring system for VEGF expression as proposed by Raica et al., as follows [9]:

SCORE 0, negative;

SCORE 1, weakly positive in less than 10% of tumour cells;

SCORE 2, weak to moderately positive in 10–50% of tumour cells;

SCORE 3, strong or moderately positive in more than 50% of tumour cells.

STATISTICAL ANALYSIS

The data were analysed using SPSS 19.0 software (IBM SPSS Inc., Chicago, IL, USA). Between-group VEGF expression levels and scores were compared using the Chi-square test. A p-value of <0.05 was considered statistically significant. The p-value was tested for two sided hypothesis.

RESULTS

VEGF expression in astrocytoma and oligodendroglioma:

VEGF expression was analysed staining with anti-VEGF antibody. While two (3.33%) cases showed no staining with the antibody, 58 cases (96%) showed positive staining for VEGF. Protein expression was detected in microvessels adjacent to endothelial cells along with the tumour cells in a few cases. VEGF expression was localised in the cytoplasm of tumour cells.

VEGF expression was analysed in various tumours. On the basis of the intensity of the staining [Table/Fig-1], two cases (3.33%) showed negative staining for VEGF, 10 cases (17.2%) showed weak staining, 17 cases (29.3%) showed weak to moderate staining, and 31 cases (53.457%) showed moderate to strong staining.

Analysis of VEGF expression across tumour grades:

Expression of VEGF was 100% in Grade II-diffuse astrocytoma (12/12), Grade II - oligodendroglioma (3/3) and Grade III tumours-anaplastic astrocytoma (2/2) Grade III – anaplastic oligodendroglioma (13/13), and 93.33% in Grade IV tumours-

Diagnosis	Negative	Weak in <10% of tumour cells	Weak to moderate in 10–50% of tumour cells	Moderate to strong in >50% of tumour cells	Total	p-value
Grade II	0	7	4	4	15	0.012
Grade III	0	1	6	8	15	
Grade IV	2	2	7	19	30	
Total	2	10	17	31	60	

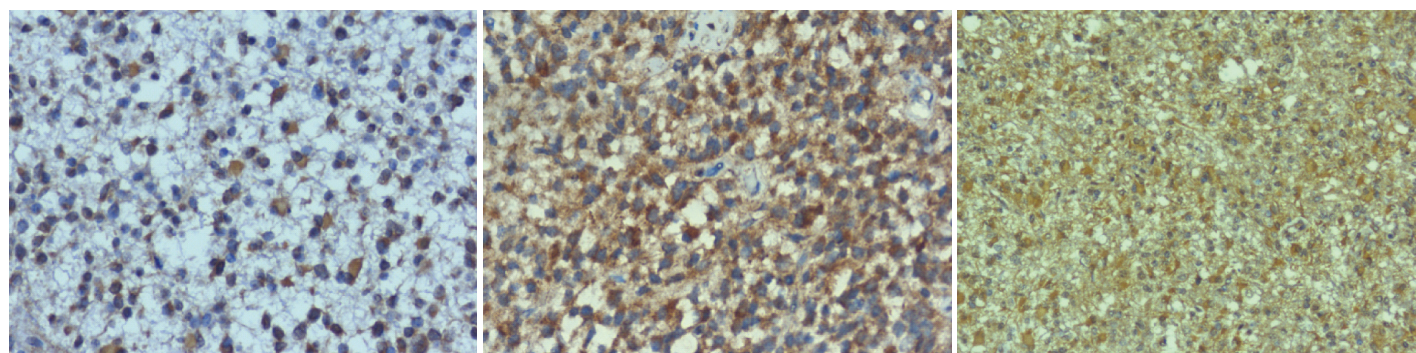
[Table/Fig-1]: Expression of VEGF across the grades. Chi-square test was used to evaluate the association between VEGF Score & Grade of the tumour

glioblastoma (28/30). The expressions of VEGF in tumours across various grades according to the VEGF scoring are shown in [Table/Fig-1]. We obtained a p-value of 0.012 (<0.05), which is significant.

Various expression details of VEGF in diffuse astrocytoma-Grade II, anaplastic astrocytoma-Grade III, glioblastoma multiforme-Grade IV have been displayed in [Table/Fig-2-4].

DISCUSSION

In the present study, we observed a higher incidence of Grade IV-glioblastoma cases than that of other grades as this is a tertiary care centre. VEGF expression positively correlated with the grades of tumour. A previous study elucidated that, VEGF expression, possibly upregulated by Phospho STAT3 (pSTAT3) in an invasion



[Table/Fig-2]: IHC- staining – VEGF cytoplasmic positivity – diffuse astrocytoma-grade II- 200x magnification – Score: 1. **[Table/Fig-3]:** IHC- staining – VEGF cytoplasmic positivity – anaplastic astrocytoma-grade III-200x magnification – Score: 2. **[Table/Fig-4]:** IHC- staining – VEGF cytoplasmic positivity –glioblastoma multiforme-grade IV-200x magnification – Score: 3.

related signalling pathway regulated by the glioma cells, may be attributed to tumour invasion [10]. The same study found that expression of VEGF was positive and highly variable in the neoplastic cells which ranged between 12%-95%. However, the positivity was assessed in glioblastoma cases only [10]. In addition, over expression of VEGF was associated with the formation of peritumoural oedema along with cell invasion in these tumours [10]. In malignant gliomas, VEGF is responsible for inducing tight junction disruptions and neovascular endothelial cell dysfunction [11–13]. In vivo studies in mouse models and glioblastoma cell lines have also shown that, transcriptional up-regulation of VEGF and its secretion is mediated by Interleukin 6 (IL-6) and dependant on STAT3 [14].

Gupta et al., have also documented significant increase in VEGF expression in high grade gliomas in comparison with the lower grades and concluded stating that increased angiogenesis contributes to astrocytoma tumourigenesis in high grade tumours [15]. Erdamar et al., have also noticed similar patterns of VEGF expression which also correlated with histological grades in malignant astrocytomas [16]. Another study demonstrated consistent expression of VEGF in tumour cells of high grade gliomas and concluded that angiogenesis is closely associated with induction of VEGF in the tumour vasculature [4].

Although, there are many molecular markers for the diagnosis of glioma, the discovery of novel therapeutic markers is essential for patient management. In this regard, VEGF serves as a novel entity for designing anti-VEGF treatment regimens; however, these are in preclinical trials in USA and other countries [17-19]. Increased VEGF expression correlates with the degree of malignancy in gliomas [20,21].

LIMITATION

The expression of VEGF in these tumours plays a significant role. However, the association of different other factors like expression of VEGF-R, p-STAT3, micro-vascular density, peritumoural oedema etc with VEGF expression has not been addressed.

CONCLUSION

VEGF plays a role in neovascularisation in tumour cells. It may have an important role in the molecular mechanisms of tumourigenesis, thus, facilitating cell migration and proliferation, endothelial cell survival, and microvascular tube formation. VEGF has proven to be useful in cancer therapy owing to its pivotal role in angiogenesis. Therefore, studies at the genetic level by using sensitive and specific molecular techniques with larger cohorts are warranted. Among the Indian population, this was the first study in which the association between VEGF and tumour progression of astrocytomas and oligodendrogliomas was examined. The association of tumour progression with other molecular markers, such as VEGF-R and p53, and cell proliferative index need to be assessed to design effective therapeutic regimens that ultimately aim at personalising therapy.

ACKNOWLEDGEMENTS

This study was funded by the Intramural Research Fund, Jawaharlal Institute of Post Graduate Medical Education and Research (JIPMER), Puducherry, India.

REFERENCES

- [1] Kargiotis O, Rao JS, Kyritsis AP. Mechanisms of angiogenesis in gliomas. *J Neurooncol.* 2006;78:281-93.
- [2] Ohgaki H. Epidemiology of brain tumours. *Methods Mol Biol.* 2009;472:323-42.
- [3] Plate KH, Breier G, Weich HA, Risau W. Vascular endothelial growth factor is a potential tumour angiogenesis factor in human gliomas in vivo. *Nature.* 1992;359:845-48.
- [4] Chan AS, Leung SY, Wong MP, Yuen ST, Cheung N, Fan YW, et al. Expression of vascular endothelial growth factor and its receptors in the anaplastic progression of astrocytoma, oligodendroglioma, and ependymoma. *Am J Surg Pathol.* 1998;22:816-26.
- [5] Katzuza J. Selected histochemical and immunocytochemical markers of biological activity of brain blood vessels. *Folia Neuropathol.* 1995;33:195-200.
- [6] Carmeliet P. Angiogenesis in life, disease and medicine. *Nature.* 2005;438:932-36.
- [7] Kaur B, Khwaja FW, Severson EA, Matheny SL, Brat DJ, Van Meir EG. Hypoxia and the hypoxia-inducible-factor pathway in glioma growth and angiogenesis. *Neuro Oncol.* 2005;7:134-53.
- [8] Louis DN, Perry A, Reifenberger G, von Deimling A, Figarella-Branger D, Cavenee WK, et al. The 2016 World Health Organization classification of tumours of the central nervous system: A summary. *Acta Neuropathol.* 2016;131:803-20.
- [9] Raica M, Cimpean AM, Anghel A. Immunohistochemical expression of vascular endothelial growth factor (VEGF) does not correlate with microvessel density in renal cell carcinoma. *Neoplasma.* 2007;54:278-84.
- [10] Wang X-F, Lin G-S, Lin Z-X, Chen Y-P, Chen Y, Zhang J-D, et al. Association of pSTAT3-VEGF signaling pathway with peritumoural oedema in newly diagnosed glioblastoma: An immunohistochemical study. *Int J Clin Exp Pathol.* 2014;7:6133-40.
- [11] Lin ZX, Yang LJ, Huang Q, Lin JH, Ren J, Chen ZB, et al. Inhibition of tumour induced oedema by antisense VEGF is mediated by suppressive vesiculovacuolar organelles (WVO) formation. *Cancer Sci.* 2008;99:2540-46.
- [12] Papadopoulos MC, Saadoun S, Davies DC, Bell BA. Emerging molecular mechanisms of brain tumour oedema. *Br J Neurosurg.* 2001;15:101-08.
- [13] Yang L, Lin Z, Huang Q, Lin J, Chen Z, Zhou L, et al. Effect of vascular endothelial growth factor on remodeling of C6 glioma tissue in vivo. *J Neurooncol.* 2011;103:33-41.
- [14] Loeffler S, Fayard B, Weis J, Weissenberger J. Interleukin-6 induces transcriptional activation of vascular endothelial growth factor (VEGF) in astrocytes in vivo and regulates VEGF promoter activity in glioblastoma cells via direct interaction between STAT3 and Sp1. *Int J Cancer.* 2005;115:202-13.
- [15] Gupta K, Radotra BD, Banerjee AK, Nijhawan R. Quantitation of angiogenesis and its correlation with vascular endothelial growth factor expression in astrocytic tumours. *Anal Quant Cytol Histol.* 2004;26:223-29.
- [16] Erdamar S, Bagci P, Oz B, Dirican A. Correlation of endothelial nitric oxide synthase and vascular endothelial growth factor expression with malignancy in patients with astrocytic tumours. *J BUON.* 2006;11:213-16.
- [17] Weathers SP, de Groot J. VEGF manipulation in glioblastoma. *Oncology (Williston Park).* 2015;29:720-27.
- [18] Holash J, Davis S, Papadopoulos N, Croll SD, Ho L, Russell M, et al. VEGF-Trap: A VEGF blocker with potent anti tumour effects. *Proc Natl Acad Sci USA.* 2002;99:11393-98.
- [19] Wachsberger PR, Burd R, Cardi C, Thakur M, Daskalakis C, Holash J, et al. VEGF trap in combination with radiotherapy improves tumour control in u87 glioblastoma. *Int J Radiat Oncol Biol Phys.* 2007;67:1526-37.
- [20] Chaudhry IH, O'Donovan DG, Brenchley PE, Reid H, Roberts IS. Vascular endothelial growth factor expression correlates with tumour grade and vascularity in gliomas. *Histopathology.* 2001;39:409-15.
- [21] Plate KH, Risau W. Angiogenesis in malignant gliomas. *Glia.* 1995;15:339-47.

PARTICULARS OF CONTRIBUTORS:

1. PhD Research Scholar, Department of Pathology, Jawaharlal Institute of Post Graduate Medical Education and Research (JIPMER), Puducherry, India.
2. Assistant Professor, Department of Pathology, Jawaharlal Institute of Post Graduate Medical Education and Research (JIPMER), Puducherry, India.
3. Associate Professor, Department of Neurosurgery, Jawaharlal Institute of Post Graduate Medical Education and Research (JIPMER), Puducherry, India.
4. Professor, Department of Pathology, Jawaharlal Institute of Post Graduate Medical Education and Research (JIPMER), Puducherry, India.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Dr. Surendra Kumar Verma,
Professor, Department of Pathology, JIPMER, Puducherry-605006, India.
E-mail: sureramyajip@gmail.com

Date of Submission: **Sep 23, 2016**

Date of Peer Review: **Oct 07, 2016**

Date of Acceptance: **Oct 29, 2016**

Date of Publishing: **Feb 01, 2017**

FINANCIAL OR OTHER COMPETING INTERESTS: As declared above.